

# PRESENCE OF CHOLINESTERASES IN *ECHINOCOCCUS GRANULOSUS* PROTOSCOLICES

GIMÉNEZ-PARDO C., ROS MORENO R.M., DE ARMAS-SERRA C. & RODRIGUEZ-CAABEIRO F.\*

## Summary :

Cholinesterases were detected in protoscolices of *Echinococcus granulosus* spectrophotometrically and electrophoretically. To characterize these activities as acetylcholinesterases or pseudocholinesterases, BW284C51 and the organophosphate anthelmintic Neguvon® were assayed as specific inhibitors of acetylcholinesterases, while Iso-OMPA was employed as specific inhibitor of pseudocholinesterases. We concluded that these cholinesterase (ChE) activities would be considered as possible targets in chemotherapy.

**KEY WORDS :** cholinesterases, *Echinococcus granulosus*, protoscolices.

## Résumé : PRÉSENCE DE CHOLINESTÉRASES AU NIVEAU DU PROTOSCOLEX D'*ECHINOCOCCUS GRANULOSUS*

Des cholinestérases ont été détectées par spectrophotométrie et électrophorèse au niveau du protoscolex d'*Echinococcus granulosus*. Les activités - acétylcholinestérase et pseudocholinestérase - ont été caractérisées en utilisant des inhibiteurs spécifiques des acétylcholinestérases : BW 284 C51 et l'anthelminthique Neguvon®. L'Iso-OMPA a été utilisé en tant qu'inhibiteur spécifique des pseudocholinestérases. Les auteurs concluent que ces activités pourraient constituer une cible en matière de chimiothérapie.

**MOTS CLÉS :** cholinestérases, *Echinococcus granulosus*, protoscolex.

There is little information about the neuromuscular system in cestode species of clinical significance (Smyth & McManus, 1989). Acetylcholine, 5-hydroxytryptamine (5-HT) and glutamate have been localized in cestodes (Webb & Eklove, 1989; Samii & Webb, 1990; Solís & De Jong, 1994; Brownlee *et al.*, 1994; Fairweather *et al.*, 1994; Terenina *et al.*, 1998) and there is a consensus that acetylcholine acts as the inhibitory neurotransmitter (Thompson & Mettrick, 1984; Sukhdeo & Mettrick, 1987), 5HT and glutamate stimulate the motility in cestodes working as the excitatory neurotransmitters (Sukhdeo & Mettrick, 1987; Webb & Eklove, 1989) and they have an activating effect on metabolism (Thompson & Mettrick, 1984). It is known that acetylthiocholine is degraded into thiocholine and acetate by means of acetylcholinesterase (AChE) activity (Ellman *et al.*, 1961).

*Echinococcus granulosus* protoscolices are waiting to become in adults upon ingestion by definitive hosts and their interest may be related to dissemination following the cyst rupture. With the finality to avoid this problem, several authors have worked with different drugs (Casado *et al.*, 1989; Pérez-Serrano *et al.*, 1997). In nematodes the ChE activities have been proposed as targets of some anthelmintics (Blackburn & Selkrik, 1992; Tekwani,

1992), in this paper, these activities in whole extracts of *E. granulosus* protoscolices were studied, and the organophosphate anthelmintic Neguvon® was assayed *in vitro* as an agent that effectively binds to AChE.

## MATERIALS AND METHODS

Protoscolices of *E. granulosus* were removed aseptically from hydatid cysts from the liver of twenty naturally infected sheep slaughtered at the municipal abattoir of Sepúlveda (Segovia, Spain). They were washed with 10mM sterile phosphate buffer solution (PBS pH 7.2) supplemented with penicillin (1 mg/ml) and dihydrostreptomycin sulphate (2 mg/ml), pooled and homogenized once at 4° C using a glass Potter-Elvehjem homogenizer. Centrifugation of the homogenates was performed 2 × at 100,000 × g for 30'. The final supernatant was considered to be the cytosolic fraction. Protein concentration was measured by a modified Lowry method (Peterson, 1983) and adjusted to 2 mg/ml. Aliquots were then used immediately or frozen at -80° C and the results from those experiences did not show differences between them. Cholinesterase (ChE) activity was determined spectrophotometrically according to Ellman *et al.*, (1961) using acetylthiocholine iodide (ATCI) or butyrylthiocholine iodide (BTCl) and electrophoretically corroborated by Karnovsky & Roots (1964) staining method with the same substrates. Briefly, 3 ml of 0.1 M PBS pH 8, 20 µl substrate, 50 µl

\* Laboratory of Parasitology, Faculty of Pharmacy, University of Alcalá, Crtra. Madrid-Barcelona Km 33.600, 28871 Alcalá de Henares, Madrid, Spain.

Correspondence: Consuelo Giménez Pardo.  
Tel.: +349 1 885 46 36 - Fax: +349 1 885 46 63.

of the sample and 100  $\mu$ l of dithiobisnitrobenzoic acid solution, (39,6 mg dithiobisnitrobenzoic acid (DTNB), 10 ml PBS pH 7 and 15 mg sodium bicarbonate), were added to each tube and the absorbance was measured at 412 nm. Moreover, it was employed an enzyme-free blank at which 20  $\mu$ l of substrate were added. The increases in OD were converted to units per litre (Ellman *et al.*, 1961). One unit is equivalent to 1  $\mu$ mol of substrate hydrolyzed per minute per mg of protein. The inhibition of the acetylcholinesterase or pseudo-cholinesterase activities, was assessed according to Ellman procedure (Ellman *et al.*, 1961) using a microplate in which 260  $\mu$ l of 0.1 M PBS buffer pH 8, 10  $\mu$ l of 10 mM dithiobisnitrobenzoic acid solution, 2.5  $\mu$ l of 75 mM substrate and 25  $\mu$ l of the sample (which were 20  $\mu$ l of protoscolices crude extracts plus 5  $\mu$ l of each inhibitor, incubated 10' at 37°C) were added to each tube and the OD increment was measured at 412 nm. Inhibitors and concentrations employed were 40,000 nM, 4,000 nM, 40 nM and 4 nM of 1,5-bis(4-allyldimethylammoniumphenyl)pentan-3-one dibromide (BW 284C51) and 50 mM, 5 mM, 0.5 mM and 0.05 mM of tetraiso-propylpyrophosphoramidate (Iso-OMPA). Experiences were performed in paralel with their respective controls in which an inhibitor-free blank was employed. These inhibitors were used since they are considered, at least for Vertebrata, as specific inhibitors of AchE (EC 3.1.1.7) and BchE (EC 3.1.1.8) respectively, and they are used in recent studies on invertebrate enzymes (Talesa *et al.*, 1997). Neguvón® (C<sub>4</sub>H<sub>8</sub>Cl<sub>3</sub>O<sub>4</sub>P) (spot-on, Bayer AG), was employed as AchE inhibitor which irreversibly inhibits the enzyme, at the following concentrations: 4  $\times$  10<sup>-3</sup> mM, 0.05 mM, 0.5 mM, 5 mM, 50 mM and 400 mM. The inhibition was calculated as the percentage of ChE activity in the controls. In every case, each test was performed in triplicate and the readings were realized ten times (one per minute).

Electrophoresis in a 6 % polyacrilamde gel without SDS (100-200  $\mu$ g protein per lane diluted 3:1 in glycerol) was performed by Laemmli method (Laemmli, 1970) for two hours, and gels were incubated in: 20 mg ATCI or BTCl dissolved in 26 ml of 0.1 M PBS pH 6 for two hours to which the following were added sequentially: 2 ml 0.1 M sodium citrate, 4 ml 30 mM CuSO<sub>4</sub>, 4 ml distilled water and 4 ml 5 mM potassium ferricyanide. Activity resulted in brown bands in the gel after two hours and it was fixed with 5 % acetic acid in deionized water.

## RESULTS AND DISCUSSION

This work has revealed that protoscolices of *E. granulosis* have cholinesterase activities (Fig. 1). These activities appeared when ATCI or BTCl were employed as substrates by Karnovsky & Roots



Fig. 1. – Cholinesterase activity in *Echinococcus granulosis* protoscolices by Karnovsky *et al.* (1964) staining method on A) Acetylthiocholine iodide (ATCI). B) Butyrlthiocholine iodide (BTCl).

(1964) staining method. Moreover, this activity was also quantified by Ellman test (Ellman *et al.*, 1961). Hydrolysis of ATCI occurs as a higher relative rate than hydrolysis of BTCl as it happens in other helminths (Vos & Dick, 1992). The degradation of ATCI was 6.98 U/mg and the BTCl hydrolysis was 4.12 U/mg, both referred to 2 mg/ml protein concentration. The use of specific inhibitors (BW 284C51 and Iso-OMPA) corroborated that protoscolices have acetylcholinesterases and pseudocholinesterases. Note the presence of AchE, as demonstrate the higher inhibition percentages of this activity by BW 284C51 (Table Ia), while Iso-OMPA (Table Ib) had no effect on cholinesterase activity when the substrate used was ATCI. When BTCl was employed as substrate, boh inhibitors showed inhibitory effect though it could be due to the low activity obtained when we used this substrate as occurs in vertebrates (Massoulié *et al.*, 1993). When Neguvón®, the specific AchE inhibitor was employed, the results have shown that ChE activities are inhibited by the drug, this inhibition being statistically significant only when we employed ATCI as substrate, while significant BTCl hydrolysis happened at high concentrations (Table IIIa, b).

Perhaps, *E. granulosis* protoscolices could release those cholinesterases in the hydatid cyst. This fact would open a new hypothesis of work since these enzymes could be able to pass through the germinal layer and come to the host-tissues destroying the host acetylcholine. Moreover, all these enzymes (acetyl and pseudocholinesterases) could be attractive targets for chemotherapeutic or immunological intervention in

**A**

Inhibitor (nM)	ChE activity (U/mg) X ± SD % inhibition	
0	6.98 ± 0.52	0
4 × 10 <sup>4</sup>	2.27 ± 0.10	68.42*
1 × 10 <sup>3</sup>	2.95 ± 0.14	58.94*
4 × 10 <sup>1</sup>	3.67 ± 0.13	47.36*
4	5 ± 0.44	28.42*

\* p ≤ 0.05.

**B**

Inhibitor (nM)	ChE activity (U/mg) X ± SD % inhibition	
0	4.12 ± 0.85	0
4 × 10 <sup>4</sup>	2 ± 0.37	76.58*
1 × 10 <sup>3</sup>	2.3 ± 0.63	73.14*
4 × 10 <sup>1</sup>	2.94 ± 0.73	66.20*
4	2.94 ± 1.3	66.20*

\* p ≤ 0.05.

Table I. – *Echinococcus granulosus* cholinesterase activity inhibited by BW. A) Substrate: ATCI. B) Substrate: BTCl. U: one unit is 1 µmol of substrate hydrolyzed per minute.

**A**

Inhibitor (mM)	ChE activity (U/mg) X ± SD	% inhibition
0	6.98 ± 0.52	0
50	4.7 ± 0.30	28.42*
5	7.7 ± 2.98	0
0.5	5.88 ± 0.77	15.78
0.05	5.58 ± 0.63	2.10

\* p ≤ 0.05.

**B**

Inhibitor (mM)	ChE activity (U/mg) X ± SD	% inhibition
0	4.12 ± 0.85	0
50	2.68 ± 0.72	69.58*
5	2.20 ± 0.67	71.75*
0.5	2.46 ± 0.47	72.08
0.05	3.30 ± 0.64	61.57*

\* p ≤ 0.05.

Table II. – *Echinococcus granulosus* cholinesterase activity inhibited by Iso-OMPA. A) Substrate: ATCI. B) Substrate: BTCl. U: one unit is 1 µmol of substrate hydrolyzed per minute.

hydatid disease as Geary *et al.* (1992) proposed in other helminth infections, and, in future, a new combination of compounds that inactivate the enzyme could allow acetylthiocholine to be accumulated causing abnormally high levels of inactivation and a total blockage of the nerve function (Thompson & Mettrick,

**A**

Inhibitor (mM)	ChE activity (U/mg) X ± SD	% inhibition
0	6.98 ± 0.52	0
400	1.50 ± 0.19	59*
50	1.69 ± 0.35	68*
5	2.30 ± 0.40	51*
0.5	3.97 ± 0.32	60*
0.05	3.52 ± 0.18	18
4 × 10 <sup>-3</sup>	3.52 ± 0.36	18

\* p ≤ 0.05.

**B**

Inhibitor (mM)	ChE activity (U/mg) X ± SD	% inhibition
0	4.12 ± 0.85	0
400	1.10 ± 0.12	35.54*
50	1.10 ± 0.14	35.54*
5	1.58 ± 0.19	0
0.5	1.47 ± 0.22	6.64*
0.05	1.25 ± 0.32	12.70
4 × 10 <sup>-3</sup>	0.7 ± 0.019	0

\* p ≤ 0.05.

Table III. – *Echinococcus granulosus* cholinesterase activity inhibited by Neguvón®. A) Substrate: ATCI. B) Substrate: BTCl. U: one unit is 1 µmol of substrate hydrolyzed per minute.

1984; Sukhdeo & Mettrick, 1987), but until now it remains open to speculation.

## ACKNOWLEDGEMENTS

We want to thank M<sup>a</sup> Angeles Urrea París for providing *Echinococcus granulosus* protoscolices and Dr Enrique Rierola Roqué (Bayer, GA) for his technical assistance with the drug. This research was supported by funds provided by Project PM96-0014 (Ministry of Education and Science, MEC, Spain).

## REFERENCES

- BLACKBURN C.C. & SELKIRK M.E. Characterization of the secretory acetylcholinesterases from adult *Nippostrongylus brasiliensis*. *Molecular and Biochemical Parasitology*, 1992, 53 (1-2), 79-88.
- BROWNLEE D.J.A., FAIRWEATHER I., JOHNSTON C.F. & ROGAN M.T. Immunohistochemical localization of serotonin (5-HT) in the nervous system of the hydatid organism *Echinococcus granulosus* (Cestoda, Cyclophyllidae). *Parasitology*, 1994, 109, 233-241.
- CASADO N., RODRÍGUEZ-CAABEIRO F., JIMÉNEZ A., CRIADO A. & DE ARMAS C. *In vitro* effects of levamisole and ivermectin

- against *Echinococcus granulosus* protoscoleces. *International Journal for Parasitology*, 1989, 19, 945-947.
- ELLMAN G.L., COURTNEY K.D., ANDRES V. & FEATHERSTONE R.M. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochemical Pharmacology*, 1961, 7, 88-95.
- FAIRWEATHER I., McMULLAN M.T., JOHNSTON C.F., ROGAN M.T. & HANNA R.E.D. Serotonergic and peptidergic nerve elements in the protoscolex of *Echinococcus granulosus* (Cestoda, Cyclophyllidae). *Parasitology Research*, 1994, 80, 649-656.
- GEARY T.G., KLEIN R.D., VANOVER L., BOWMAN J.W. & THOMPSON D.P. The nervous systems of helminths as targets for drugs. *Journal for Parasitology*, 1992, 78 (2), 215-230.
- KARNOVSKY M.J. & ROOTS L. A "direct-coloring" thiocoline method for cholinesterases. *Journal of Histochemical Cytochemistry*, 1964, 12, 219-221.
- LAEMMLI U.K. Cleavage of structural proteins during the assembly of the head of bacteriophage T<sub>4</sub>. *Nature*, 1970, 227, 680-685.
- MASSOULIÉ J., PEZZEMENTI L., BON S., KREJCI E. & VALLETTE F.M. Molecular and cellular biology of cholinesterases. *Progress in Neurobiology*, 1993, 41, 31-91.
- PÉREZ-SERRANO J., DENEGRI G., CASADO N. & RODRIGUEZ-CAABEIRO F. *In vivo* effect of oral albendazole and albendazole sulphoxide on development of secondary echinococcosis in mice. *International Journal for Parasitology*, 1997, 27 (11), 1341-1345.
- PETERSON G.L. Determination of total protein. *Methods in Enzymology*, 1983, 91, 95-119.
- SAMII S.I. & WEBB R.A. Acetylcholine-like immunoreactivity in the cestode *Hymenolepis diminuta*. *Brain Research*, 1990, 513, 161-163.
- SMYTH J.D. & MCMANUS P. The Physiology and Biochemistry of Cestodes. Press Syndicate of the University of Cambridge. The Pitt Building, Trumpington Street, Cambridge. CB2 1RP, 32 East 57th Street, New York NY 10022, USA, 1989, 398 p.
- SOLÍS-SOTO J.M. & DE JONG-BRINK M. Immunocytochemical study on biologically active neurosubstances in daughter sporocysts and cercariae of *Trichobilharzia ocellata* and *Schistosoma mansoni*. *Parasitology*, 1994, 108 (3), 301-311.
- SUKHDEO M.V.K. & METTRICK D.F. Parasite behaviour: Understanding plathyhelminth responses. *Advances in Parasitology*, 1987, 26, 73-144.
- TALESA V., ROMANI R., GRAUSO M., ROSI G. & GIOVANINNI E. Expression of a single dimeric membrane-bound acetylcholinesterase in *Parascaris equorum*. *Parasitology*, 1987, 115, 53-660.
- TEKWANI B.L. Secretory cholinesterase of *Ancylostoma ceylanicum*: effect of tubulin binding agents and benzimidazole anthelmintics. *Life Sciences*, 1992, 50, 747-752.
- TERENINA N.V., KOSOVATOVA L.A., GERASIMOVA E.I. & SHA-LAEVA N.M. The effect of anthelmintics on the serotonin content of cestodes. *Medidsinkaia Parazitologiya y Parazitarnye Bolezni*, 1998, Jan-Mar (1), 10-15.
- THOMPSON C.S. & METTRICK D.F. Neuromuscular physiology of *Hymenolepis diminuta* and *Hymenolepis microstomia* (Cestoda). *Parasitology*, 1984, 89, 567-578.
- VOS T. & DICK T.A. Characterization of cholinesterases from the parasitic nematode *Trichinella spiralis*. *Components in Biochemical Physiology*, 1992, 103C (1), 129-134.
- WEBB R.A. & EKLOVE H. Demonstration of intense glutamate-like immunoreactivity in the longitudinal nerve cords of the cestode *Hymenolepis diminuta*. *Parasitology Research*, 1989, 75, 545-548.

Reçu le 4 octobre 1999  
 Accepté le 30 décembre 1999